

RESEARCH ARTICLE

Open Access

HOXA13 and *HOXD13* expression during development of the syndactylous digits in the marsupial *Macropus eugenii*

Keng Yih Chew^{1,2}, Hongshi Yu^{1,2}, Andrew J Pask^{1,2,3}, Geoffrey Shaw^{1,2} and Marilyn B Renfree^{1,2*}

Abstract

Background: Kangaroos and wallabies have specialised limbs that allow for their hopping mode of locomotion. The hindlimbs differentiate much later in development but become much larger than the forelimbs. The hindlimb autopod has only four digits, the fourth of which is greatly elongated, while digits two and three are syndactylous. We investigated the expression of two genes, *HOXA13* and *HOXD13*, that are crucial for digit patterning in mice during formation of the limbs of the tammar wallaby.

Results: We describe the development of the tammar limbs at key stages before birth. There was marked heterochrony and the hindlimb developed more slowly than the forelimb. Both tammar *HOXA13* and *HOXD13* have two exons as in humans, mice and chickens. *HOXA13* had an early and distal mRNA distribution in the tammar limb bud as in the mouse, but forelimb expression preceded that in the hindlimb. *HOXD13* mRNA was expressed earlier in the forelimb than the hindlimb and was predominantly detected in the interdigital tissues of the forelimb. In contrast, the hindlimb had a more restricted expression pattern that appeared to be expressed at discrete points at both posterior and anterior margins of the limb bud, and was unlike expression seen in the mouse and the chicken.

Conclusions: This is the first examination of *HOXA* and *HOXD* gene expression in a marsupial. The gene structure and predicted proteins were highly conserved with their eutherian orthologues. Interestingly, despite the morphological differences in hindlimb patterning, there were no modifications to the polyalanine tract of either *HOXA13* or *HOXD13* when compared to those of the mouse and bat but there was a marked difference between the tammar and the other mammals in the region of the first polyserine tract of *HOXD13*. There were also altered expression domains for both genes in the developing tammar limbs compared to the chicken and mouse. Together these findings suggest that the timing of *HOX* gene expression may contribute to the heterochrony of the forelimb and hindlimb and that alteration to *HOX* domains may influence phenotypic differences that lead to the development of marsupial syndactylous digits.

Background

The limbs are highly variable structures between different mammalian and vertebrate species [1] enabling them to adapt and exploit new habitats. The vertebrate limb has served as a key model for understanding the signalling pathways controlling patterning and morphogenesis [2]. Limb patterning genes and pathways have

been well described in mice and chickens, but very few other model animals have been examined [1,3,4].

The tammar wallaby, *Macropus eugenii*, like all macropodid marsupials, has large hindlimbs specially adapted for hopping. The digits on the hindlimb are highly modified: digit 1 is never present, but digits 2 and 3 are fused and there is an elongated digit 4 [5]. The tammar delivers an altricial young which climbs to the pouch using its relatively well developed forelimbs, but the hindlimbs are not yet functional and are essentially fetal. After birth, this situation changes and the hindlimb growth and development rapidly overtakes

* Correspondence: m.renfree@unimelb.edu.au

¹ARC Centre of Excellence in Kangaroo Genomics, The University of Melbourne, Melbourne, Victoria, 3010, Australia
Full list of author information is available at the end of the article

that of the forelimb during early pouch life. Despite this difference in timing, the tammar hindlimb autopod is specified before birth and the early formation of the syndactylous hindlimb digits is already initiated. However, the gene(s) regulating this process are as yet unknown.

Patterning of the vertebrate limb is coordinated by morphogens secreted across three different axes in the early limb bud; proximal distal (PD), dorsal ventral (DV) and anterior posterior (AP) [reviewed by [2]]. These morphogenic gradients dictate the formation of the stylopod (shoulder), zeugopodium (radius and ulna) and autopodium (hand and digits) [reviewed by [6]]. Fibroblast growth factor 8 (FGF8) expressed in the Apical ectodermal ridge (AER) controls a positive feedback signal that dictates PD outgrowth [7]. Bone morphogenetic protein 4 (BMP4) is a key regulator of interdigital webbing and dorsal ventral polarity [8] while sonic hedgehog (SHH) secreted from zone of polarising activity (ZPA) (a signalling centre in the posterior region of the limb bud) acts via a negative feedback loop with BMP4 to determine anterior posterior patterning and digit identity [9],

Interestingly, many of these genes and proteins also play conserved roles in the development of another appendage, the phallus [10,11]. SHH is secreted from the urethral epithelium of the phallus and regulates patterning much like the ZPA in the limb bud [11,12]. BMP4 is expressed in the phallus in the distal region near the urethral epithelium and interacts with SHH [reviewed by [11]]. Although these appendages are phenotypically different, the main signalling pathways remain the same [11] and *Homeobox* (HOX) genes underpin their regulation.

The HOX genes are crucial regulators of embryonic development and the phenotypic differences in the vertebrate body plan [reviewed by [13]] and are responsible for patterning the limb [reviewed by [12,14,15]]. *Homeobox A13* (*HOXA13*) and *Homeobox D13* (*HOXD13*) are essential for formation of the autopod and digit patterning in the mouse [16]. *HOXA13* mutants have fused digits and there is no digit 1 (the most anterior digit) [16]. *HOXD13* mutations result in fusion of digits 3 and 4 and a localised delay in autopod ossification [16]. *HOXA13* and *HOXD13* double heterozygous mutants have more severe limb and genital phenotypes compared to either individual gene mutation [16].

Similarly in humans, mutations in *HOXA13* and *HOXD13* result in limb and genital malformations such as synpolydactyly, polydactyly and hypospadias [17-19]. Many of these mutations are due to the expansion or reduction of a polyalanine tract in these two genes [20,21], changing the biochemical conformation [22]. Interestingly, the chicken and zebrafish have shorter polyalanine tracts and both possess highly modified

appendages [23]. It is still not clear how the *HOX* genes are regulated [14] but human mutations such as Townes-Brocks [24] suggests that the transcription factors *SALL1* and *SALL3* influence the SHH and HOX pathways [25]. Double null *SALL1/SALL3* mutants have lost digit 1 and have fused digits 2 and 3, much like the *HOXA13/HOXD13* double heterozygous mutants [25].

The bat *Carollia perspicillata*, like all chiropterans, has a highly specialised forelimb that evolved to enable flight [4]. There is an expanded and posteriorly-shifted *HOXD13* expression in the forelimbs compared to expression in the mouse [3,26]. In addition, bats have developed a mechanism of interdigital retention involving the regulation of *BMP4* and *FGF8* through *Gremlin* (*GREM1*) [8]. *SHH* has a second wave of expression in the bat forelimb [27] and may reinitiate the loop between FGF and SHH to retain the interdigital webbing of the bat and elongate the forelimb digits [27]. In contrast, the macropodid marsupials have elongated and expanded their hindlimb digits, so a comparison of expression profiles for key limb patterning factors in the tammar may help to define the mechanisms underlying vertebrate limb diversity.

To date there is only limited information about the molecular control of marsupial limb development. The grey short-tailed opossum *Monodelphis domestica* has precocious forelimb development (as is also seen in the tammar) [1,28] that facilitates the crawl from the birth canal to the mammary glands, but the hindlimb development lags slightly behind [29,30]. This heterochronic development of the forelimbs is reflected by the timing of gene expression. Paired-like homeodomain transcription factor 1 (*PITX1*) expression an upstream inducer of T-box 4 (*TBX4*) is expressed late in opossum development relative to the mouse [29]. *TBX4* and *T-box 5* (*TBX5*) are markers of hindlimb and forelimb position respectively and in the opossum are expressed relatively early in development, indicating that the opossum forelimb field arises relatively earlier than in the mouse and earlier than the opossum hindlimb [29,31]. However, unlike the tammar, the opossum limbs are not especially modified and have all 5 digits [32].

We therefore focussed on the spatio-temporal changes of two genes, *HOXA13* and *HOXD13*, known to be essential for digit development in the mouse and chicken, to investigate the possible role of *HOX* genes during digit and limb development in the tammar and to determine whether differential patterning accounts for their unique digit modifications.

Methods

Animals

Tammar embryos were collected from a wild population on Kangaroo Island, South Australia. Morphological

observations and measurements were made on a collection of 28 embryo and fetus samples. Age was estimated on embryo images and developmental growth curves given in [33-35]. Due to different growth rates and a poor correlation between age and stage in individual animals, there is some variation of limb phenotypes relative to other characters. Age of some specimens have therefore been estimated to within a half day. Fetal samples were collected from day 18 to day 25 of the 26.5 day pregnancy (n = 3 at each stage for *in situ* and PCR except for a single d18 embryo). Tissues for sequencing were obtained from the same individuals as used for the PCR analyses.

Fetal limbs were dissected from the main body trunk and snap frozen for RT-PCR or fixed whole in 4% paraformaldehyde. Three additional animals (2 fetuses at day 23 and 25 of pregnancy and three pouch young at day 3, 120 and 150 post-partum (pp) as well as an adult) were used for cartilage and bone imaging. All sampling techniques and collection of tissues conformed to Australian National Health and Medical Research Council (2004) guidelines and were approved by The University of Melbourne Animal Experimentation & Ethics Committees.

Extraction and isolation of tammar *HOXA13* and *HOXD13*

Partial sequences of tammar *HOXA13* and *HOXD13* were identified from the NCBI tammar genome trace archives, and the full sequences were obtained by BAC screening and BAC DNA 454 shotgun sequencing (H Yu, Z-P Feng, R O'Neill, Y Hu, A Pask, D Carone, J Lindsay, G Shaw, S Frankenberg, AT Papenfuss and MB Renfree, unpublished data). Primers for RT-PCR and whole mount *in situ* probes were designed to span an intron and exon boundary based on cloned BAC DNA sequence (Table 1). Total RNA was extracted from tammar fetal limbs and cDNA synthesis was performed according to the manufacturer's protocol (Invitrogen, NSW, Australia). To examine *HOXA13* and *HOXD13* expression in developing limbs and to make *in situ* templates, RT-PCR was performed using the following conditions: 35 cycles of 30s, 95°C; 30s, 58°C; 60s, 72°C, in a 25 µl reaction with GoTaq Green Master Mix (Promega, Wisconsin, USA). The RNA control was a pooled template before reverse transcribed into cDNA of forelimb or hindlimb.

HOXA13 and *HOXD13* protein sequences of human, mouse, opossum, platypus, chicken and zebrafish were retrieved from GenBank (<http://www.ncbi.nlm.nih.gov/>) (Additional file 1, Table S1). The protein sequences were aligned with MUSCLE program [36]. The phylogenetic tree was constructed by neighbour-join method with default parameter of program MEGA5 [37]. The second structure of *HOXD13* N-terminal was predicted using PHYRE <http://www.sbg.bio.ic.ac.uk/~phyre/> [38] and the second structure is the consensus structure predicted by pispred, jnet and sspro.

Whole mount *in situ* hybridisation

Whole mount *in situ* hybridisation was performed as described previously [38]. Embryos were fixed in 4% paraformaldehyde at 4°C overnight, washed and stored in 100% methanol at -20°C until analysis. Limbs were dissected from the fetal bodies and rehydrated through a graded methanol series (10 mins each). Fetuses were washed in PBS with 0.1% triton X-100 (PBTX) (3 × 10 mins) before proteinase K treatment 10 µg/ml (30-60 minutes). Fetal limbs were washed in PBTX for (2 × 5) minutes and fixed with 0.2% glutaraldehyde/4% PFA in PBTX for 20 minutes on a rocker. Embryos are then incubated in pre-hybridisation mix at 65°C overnight before 1 µg of a cRNA probe synthesized with T7 or SP6 RNA polymerase (Promega, Australia) was added and incubated overnight at 65°C. Using saline-sodium citrate (ssc)/0.5% 3-[(3-cholamidopropyl) dimethylamino]-1-propanesulfonate (CHAPS), tissues were washed and blocked for 2-3 hours and incubated overnight in alkaline phosphate-conjugated DIG antibody (Roche Diagnostics Corporation, Indianapolis, IN). The specimens were washed in Tris-buffered saline with triton X-100 (4 × 60 mins) before colour development. All specimens were incubated in nitro-blue tetrazolium and 5-bromo-4-chloro-3'-indolylphosphate (NBT/BCIP) for a minimum of 2 hours. Photographs were taken using an Olympus dp25 camera mounted on an Olympus (ZX-9) dissection microscope.

Bone and cartilage staining

Alcian blue staining (fetal stages)

Alcian blue staining of fetal stages was carried out as described by [39] with the following modifications.

Table 1 Primers used in this study (5' to 3')

Gene	Forward	Reverse	Amplicon size (BP)
<i>HOXA13</i> (WISH)	CTACTTCGGCAGCGGTTA	TGTCGCTTGAGTTTGTGA	676
<i>HOXD13</i> (WISH)	GTTTCATCCTCTTCTCCTC	AATGGCGTATTCGTTCTC	656
<i>HOXA13</i> (PCR)	ACCTCTGGAAGTCCACTCTG	CTTGGTGTAAGGCACTCGTT	95
<i>HOXD13</i> (PCR)	CTGGACTCTGGCGAATGG	GGTTTGTGGCTGCGGATA	251
M13puc (sequencing)	CCCAGTCACGACGTTGTAAAACG	AGCGGATAACAATTTACACAGG	

Specimens were initially fixed in 4% paraformaldehyde (PFA) overnight and then washed and stored in 70% ethanol before staining. Specimens were then washed in a 1:1 mixture of ethanol and NaOH before staining.

Alcian blue and alizarin red staining of postnatal bone

All specimens were initially fixed in 4% PFA overnight and then washed and stored in 70% ethanol before staining. As much skin and fat as possible was carefully removed before placing the eviscerated specimen in alcian blue solution (0.05% Alcian Blue 8 GX (Prositech; Queensland, Australia) in 95% ethanol, 5% acetic acid) overnight. The following day the specimens are washed in 95% ethanol overnight and then incubated in 2% KOH for 1-2 days. Specimens are then placed in Alizarin red solution (0.1% alizarin red in 1% KOH) for 2 days. Specimens are then cleared in 1% KOH, 20% glycerol for at least 2 days or until clear before photography.

Results

Tammar limb development

The forelimbs of adult tammar are similar to those of the mouse and have five digits making a hand that is well adapted to grasping (Figure 1). However, the tammar hindlimb has only four digits. Tammar digit one is never present at any stage of development whilst digits two and three are fused (syndactyly). The fourth digit is greatly elongated and the fifth digit is reduced. The syndactylous form of the limb is specified between day 23 and day 24 of pregnancy (Figure 2). There are differences in the developmental timing of fore- and hindlimbs.

At day 19 of pregnancy the tammar forelimb formed initially as a bud whilst the hindlimb was a ridge less than half of the forelimb bud size. The forelimb transitioned from a club-shaped bud structure to a paddle between day 19 and day 20 but digital rays were only visible on the dorsal surface towards the end of this stage. The hindlimb developed into a pronounced outgrowth that resembled a bud by day 20-2 of the 26.5 day pregnancy (Figure 2A and 2B).

By day 22, the digital rays of the forelimb were well defined from the mesenchyme with interdigital webbing (Figure 2C). The digits were at an early stage of separation but the marsupial epitrichial claws that are used to assist the climb to the pouch were not yet developed. In contrast, the hindlimb was an asymmetrical bud beginning to form digital condensations.

Epitrichial claws formed at day 24, and the digits had separated, extending forward but were uncurled. The hindlimb had rays formed by digital primordia with a single enlarged digit four that eventually would form the asymmetrical hindlimb. At the most anterior point, digits two and three were distinct but shorter in length

than digit 4. This was the first sign of the developing syndactylous digits (Figure 2D).

At day 25, one day before birth, the forelimb had a well-formed elbow and keratinised epitrichial claws (Figure 2E). These claws were curved towards the ventral axis to form a fist and the living fetus repeatedly opened and closed the fist as well as moved the arms in a swimming motion. In contrast, the hindlimb was immobile and remained at right angles to the body. The hindlimb interdigital webbing had regressed and distinct digits were visible but the digits had not separated from each other (Table 2). However, the limbs had no ossification centres and contained only cartilaginous elements (Figure 3A and 3B) as detected using alcian blue and alizarin red staining, and by day 3 post-partum only the radius and ulna had begun to ossify (data not shown).

Conservation and evolution of gene structure of *HOXA13* and *HOXD13*

Both *HOXA13* and *HOXD13* have two exons that encoded 393 and 341 amino acids respectively. The HOX homeodomain was highly conserved in both *HOXA13* and *HOXD13* as in human, mouse, opossum, tammar, bat and chicken protein alignments (Figure 4A and 4B).

Tammar *HOXA13* polyalanine tracts were highly conserved with those of human, bat and mouse, but not with the chicken. In contrast, the single tammar *HOXD13* polyalanine tract was highly conserved in all species. However, the single opossum and tammar *HOXD13* polyserine tract was conserved with the second of the two polyserine tracts that are present in human, mouse and bat (Figure 4B). Interestingly, there was a marked difference between the tammar and all the other mammals examined in the region of the first eutherian polyserine tract of *HOXD13*. It also differed markedly from this region in the opossum. Another region of difference was immediately downstream of the eutherian polyserine tract in which the amino acids GQCR are conserved in all species, including the opossum, but they are absent in the tammar (Figure 4B). In order to further compare the N-terminal region of *HOXD13* in the human, opossum and tammar, the secondary structure was predicted and showed that the sequence in the tammar in the region of the first polyserine tract of the eutherians produced a 13- α -helix structure instead of the 3- α -helix structure that is present in both the human and opossum (Figure 4C). The conserved polyalanine tract formed a predicted α -helix structure but a third α -helix was also present in both opossum and tammar but absent from human (Figure 4C).

***HOXA13* and *HOXD13* expression in the tammar autopod**

RT-PCR was performed as an initial examination of *HOXA13* and *HOXD13*. Both genes were detected from

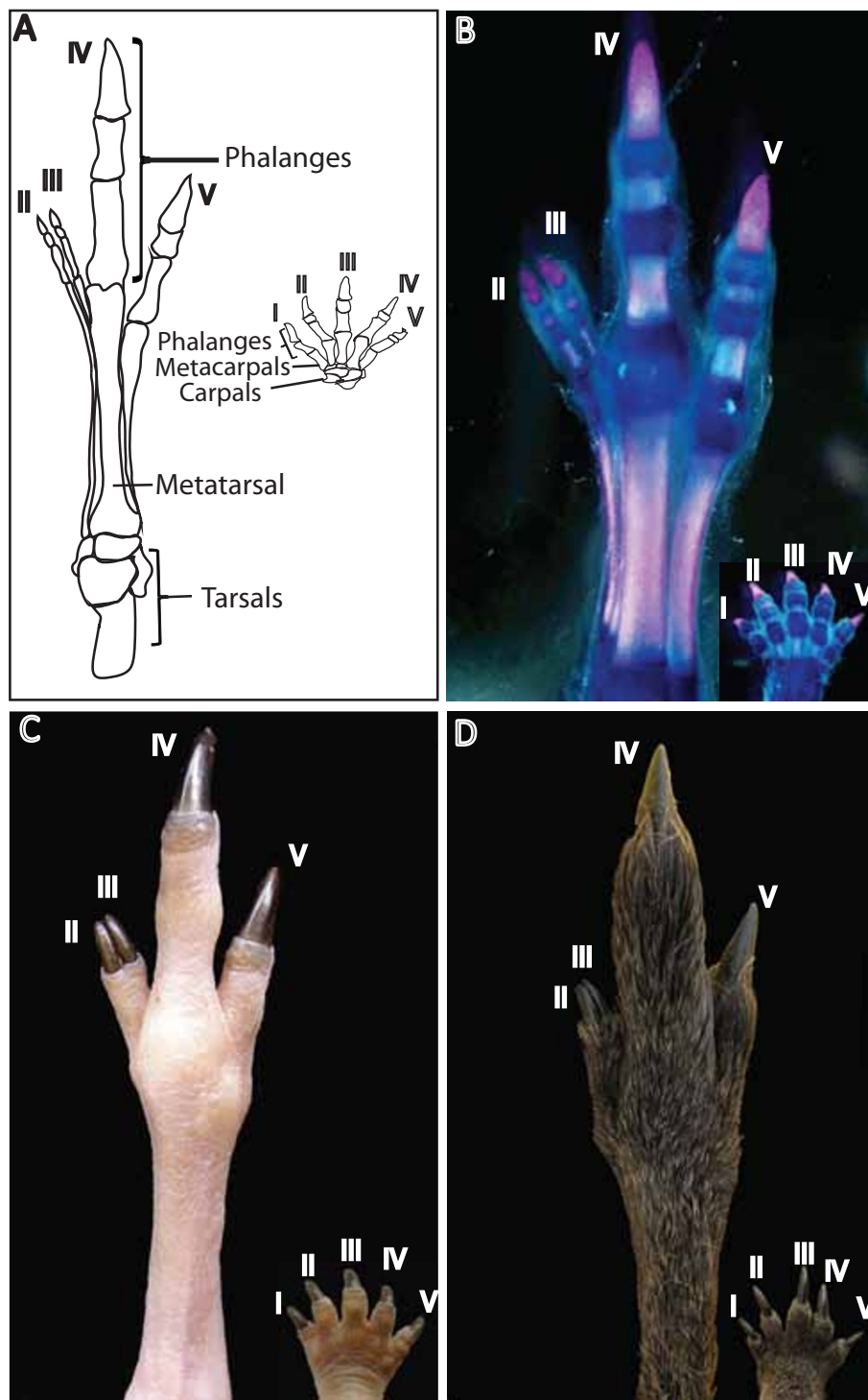
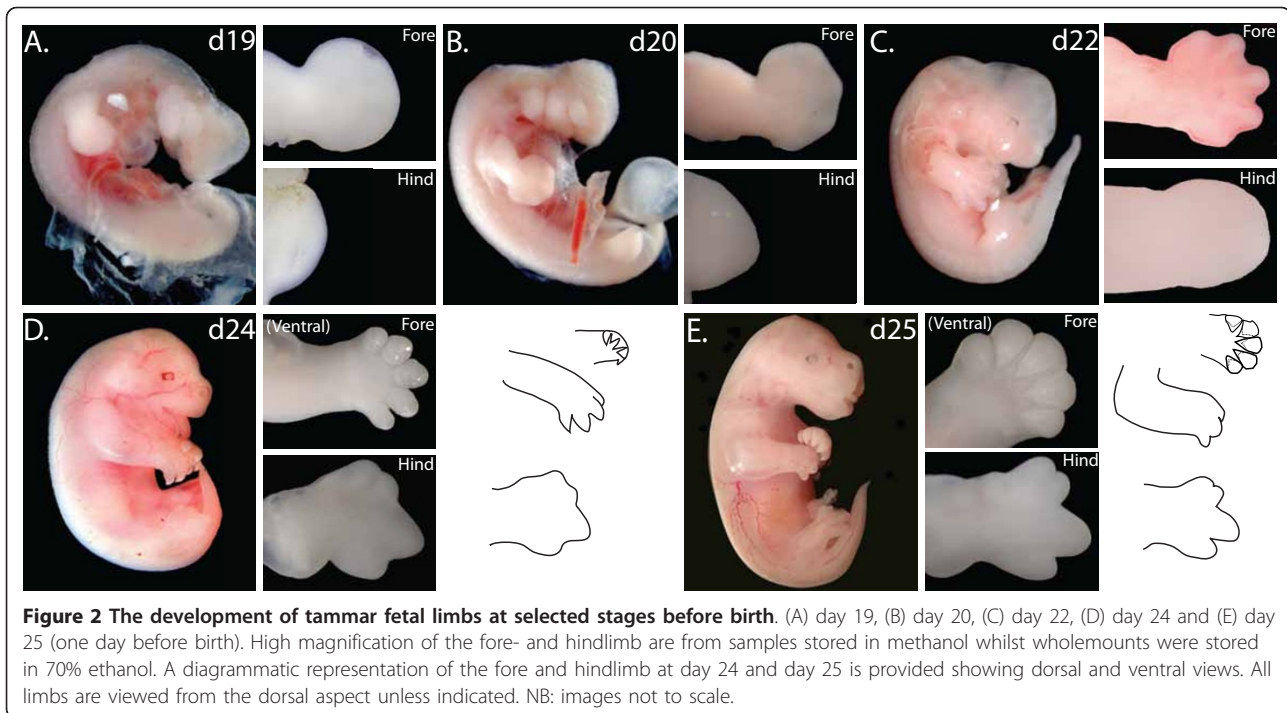


Figure 1 The morphology of the tammar wallaby autopod (A) Diagram of the macropodid forelimb and hindlimb bones (Adapted from [53]). (B) A day 150 post-partum (pp) forelimb (inset) and hindlimb stained for bone and cartilage with alcian blue and counter stained with alizarin red. (C) A day 120 pp forelimb (inset) and hindlimb. (D) An adult tammar forelimb (inset) and hindlimb. In the forelimb, all five digits were present. In contrast the most anterior digits of the hindlimb were the syndactylous digits two and three. Digits are numbered with Roman numerals.



day 21 to day 25 of pregnancy in both the fore- and hindlimb (Figure 5). *HOXA13* and *HOXD13* expression during tammar limb development was also examined using wholemount *in situ* hybridisation (Figure 6). *HOXA13* mRNA had an early and transient expression in the tammar autopod. Expression was first detected at day 18.5 in the distal margins of the forelimb paddle and extended from the anterior to the posterior margins. At the same stage there was no detectable expression in the hindlimb. Later at day 21, *HOXA13* mRNA was restricted to the interdigital regions of the forelimb. The only detectable *HOXA13* expression in the hindlimb was in the distal region of the hindlimb bud at day 21. In both the forelimb and hindlimb, *HOXA13* transcripts was detected at day 24 and day 25.

HOXD13 expression was expressed distally in the day 18 and day 19 forelimb showing a similar expression pattern to the staining seen in *HOXA13* at an equivalent stage (Figure 6B). There was no expression detected in the hindlimb at this stage. Two days later at day 21.5 *HOXD13* was strongly expressed in the forelimb interdigital region and extended from the start of the digital condensations to the distal tip of the paddle. Unlike the forelimb, there was no detectable expression in the hindlimb bud.

At day 23.5, *HOXD13* regressed towards the proximal boundary away from the tips of autopod. The first expression was detected in the hindlimb on this day of pregnancy and was expressed in the proximal and distal regions of the bud. At day 24, when the forelimb digits

Table 2 Summary of limb development before birth in the tammar fetus

STAGE	FORELIMB	HINDLIMB
Day 18	Bud has formed with no defined shape as yet	Thickening of the hindlimb has occurred with a small protrusion as the beginnings of a bud
Day 19-21	Paddle like structure with the regions of the future digital rays beginning to form.	Elongation of the bud has occurred with no distinct structure
Day 22-23	Digital rays more pronounced and the interdigital tissue has begun to regress	The bud has formed into a flattened arrow-head like structure with the beginnings of the digital condensations. The pointed edge of the paddle appears to be the presumptive 4th digit
Day 23-24	Interdigital webbing has regressed. Epitrichial claws are present and fingers are open but not clenched. The distinct protrusion of the future elbow joint is beginning to form	The digital rays have begun to form and a distinct separation of the hindlimb digits begins. The interdigital webbing is reduced and an asymmetrical shape has started to form
Day 25	A well-defined forelimb with claws present with digits in a clenched position. The future elbow joint has become more pronounced	The interdigital webbing has regressed and all four digits are distinct. The 4th digit has become more pronounced and has an asymmetrical shape.

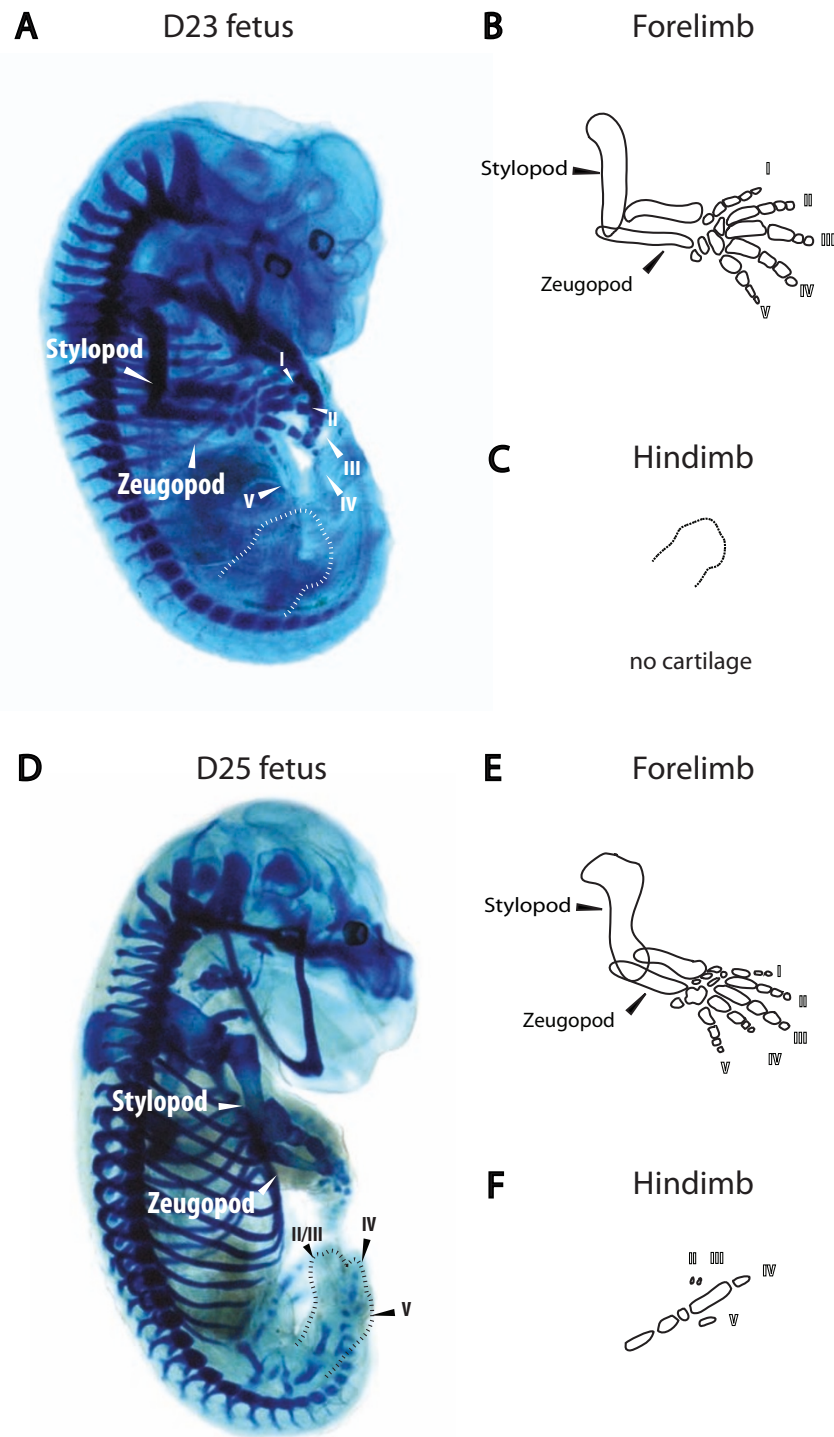
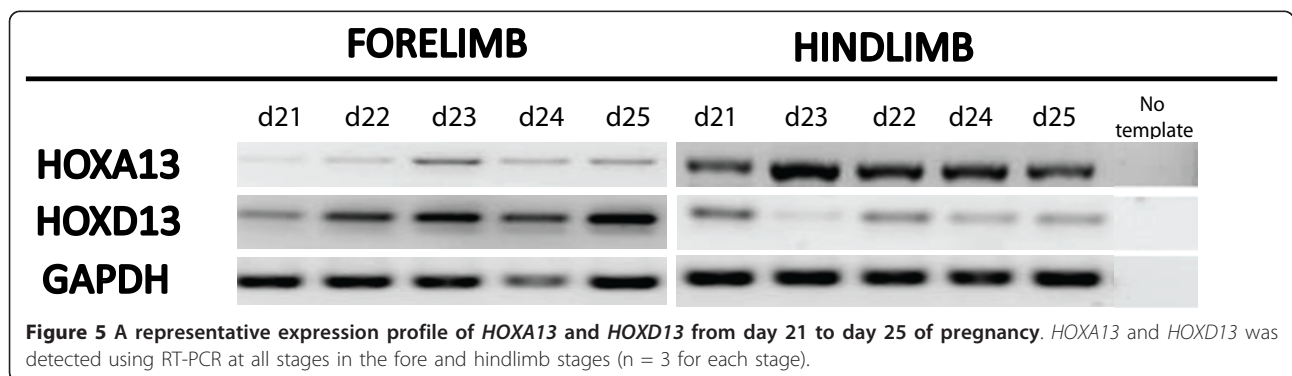
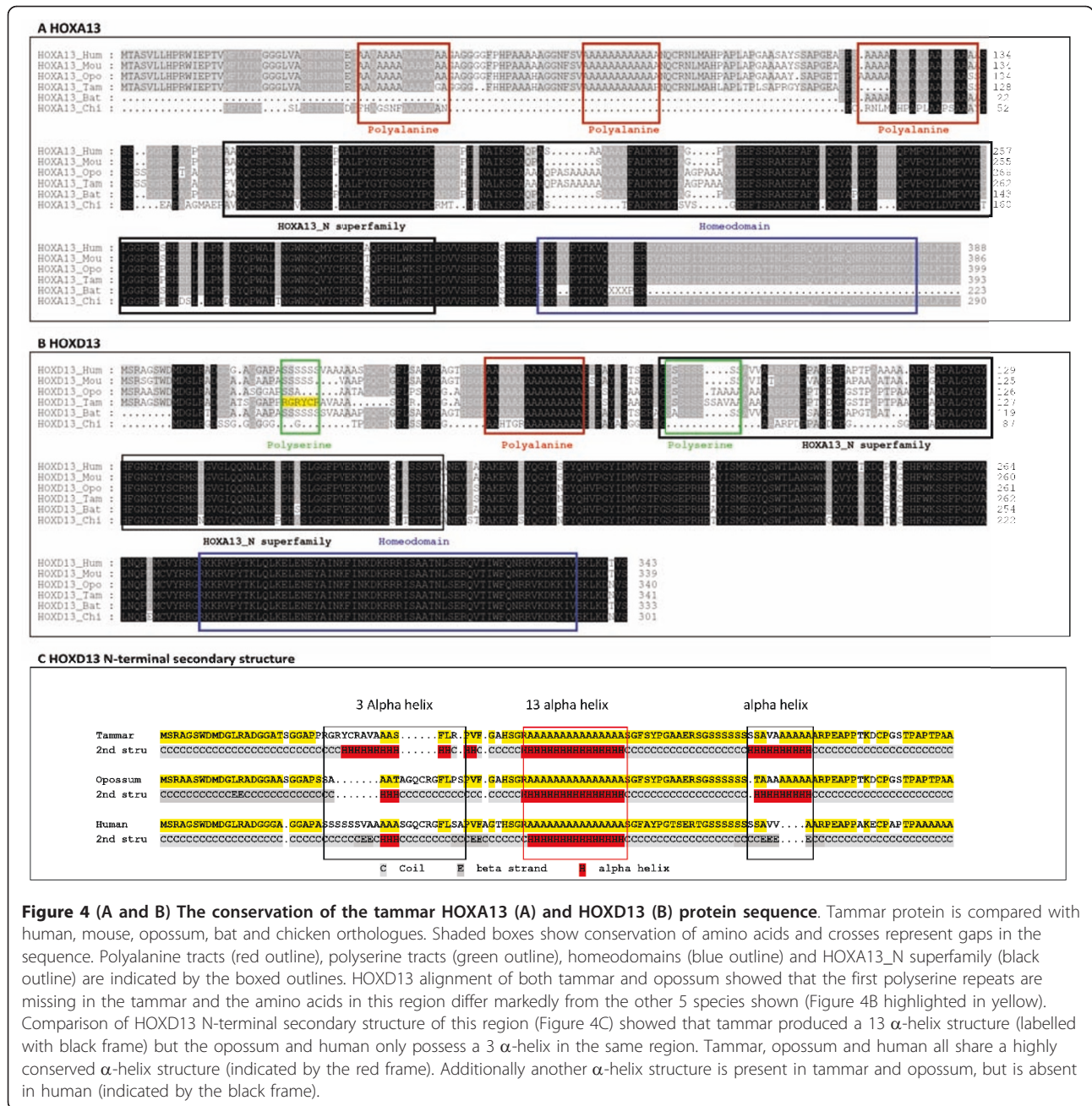
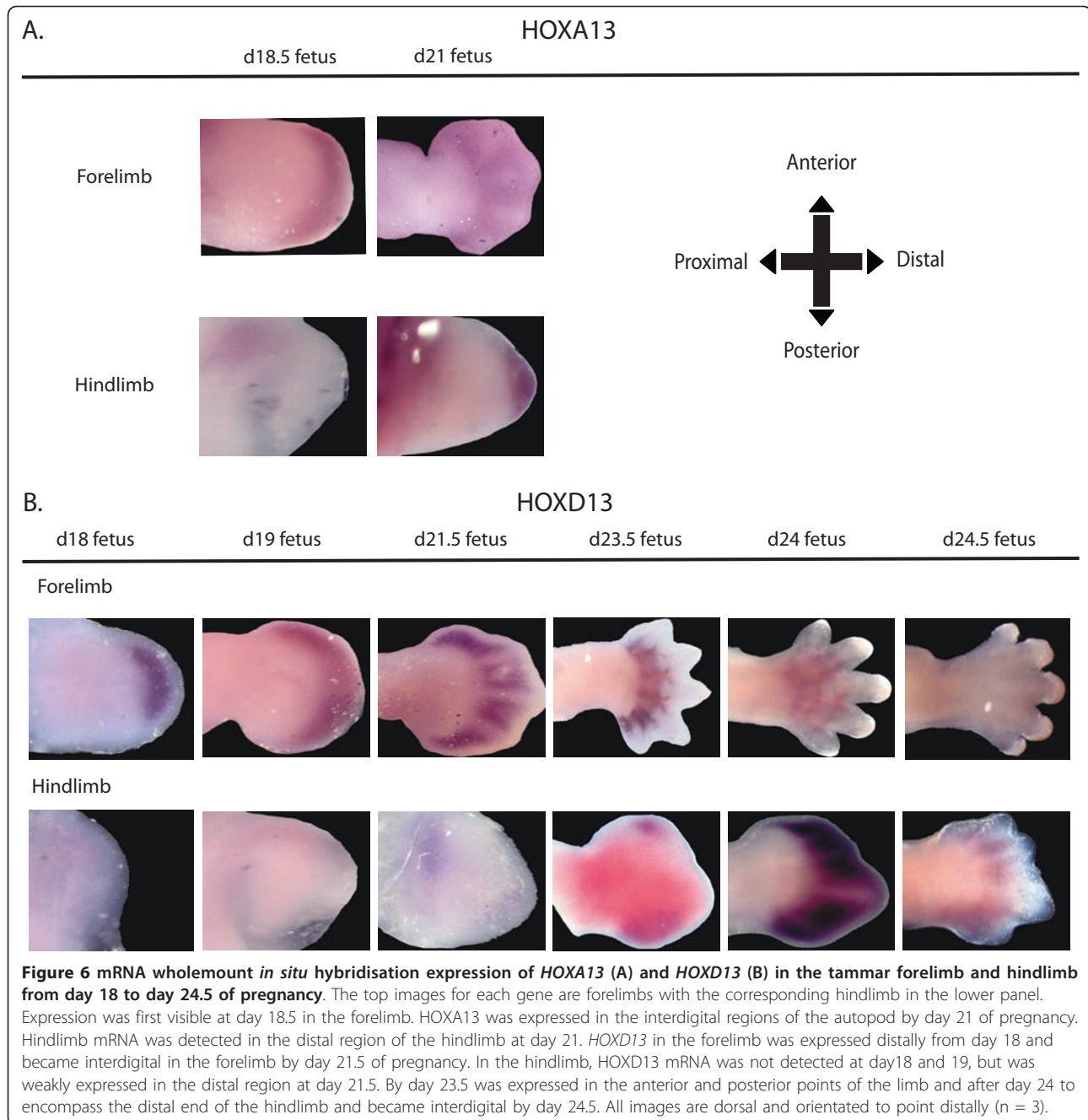


Figure 3 The development of bone and cartilage in the tammar limb. Alcian blue staining of (A) day 23 fetus and (D) day 25 fetus. A diagrammatic representation of alcian blue stained cartilage elements in a day 23 fetus (B) forelimb, (C) hindlimb and day 25 fetus (E) forelimb and (F) hindlimb. Staining show the early skeletal element deposited as cartilage was clearly visible in the forelimb before birth but not in the hindlimb. By day 25 of pregnancy, the forelimb was well formed and the hindlimb shows early cartilage elements. Digits are numbered with Roman numerals.

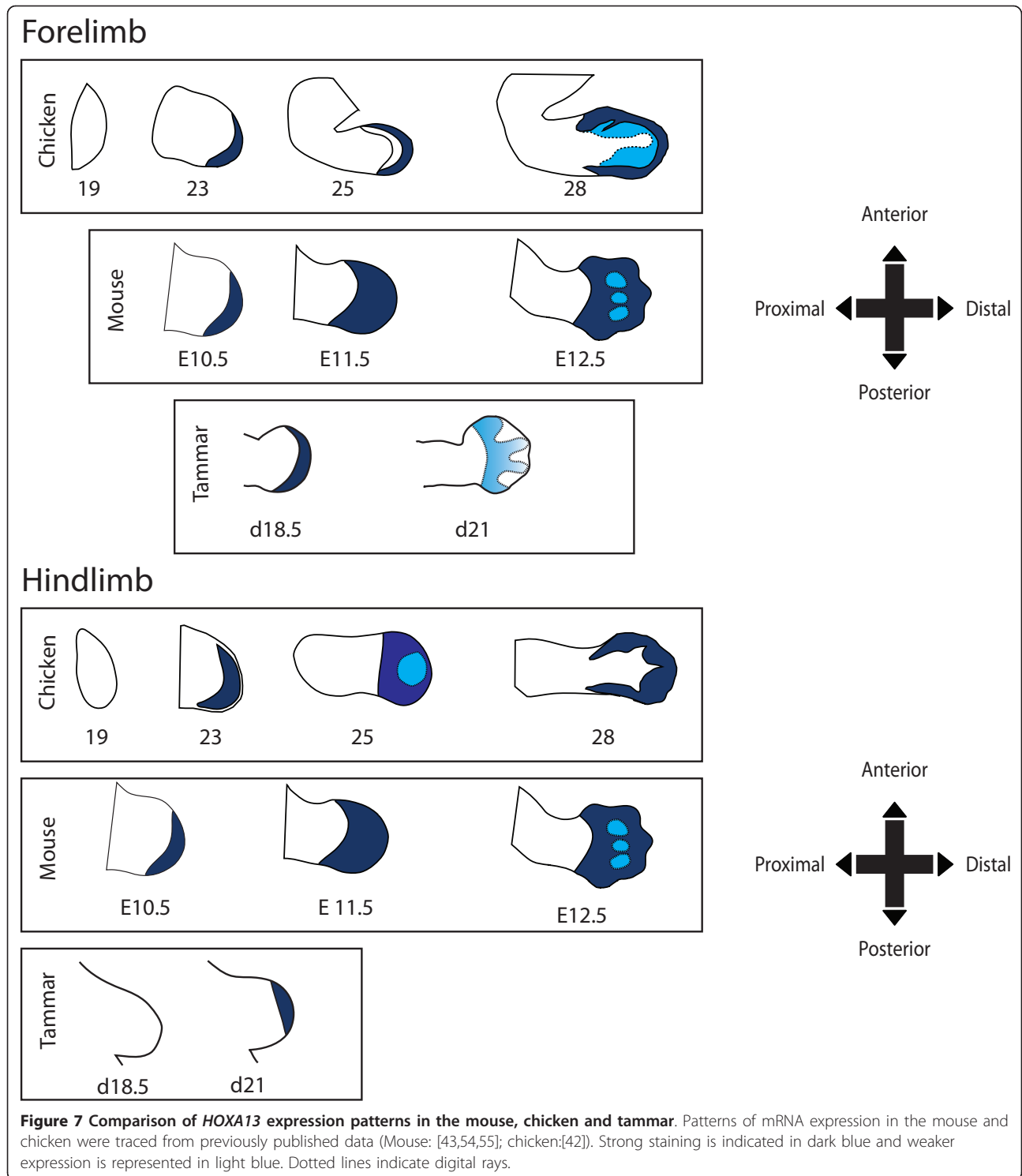




were clearly defined, the expression between the digits was weaker compared to day 23.5. Whilst expression was strongest in the hindlimb at this stage, *HOXD13* was only expressed in the interdigital regions. By day 24.5, one day before birth, there was only weak hindlimb expression and no detectable expression in the forelimb. However, in these older and larger specimens this reduced expression may have been due to poor probe penetration.

Tammar, mouse and chicken *HOXA13* expression

The mouse and chicken *HOXA13* expression pattern summarised diagrammatically from previous studies [40,41] were compared with that of the tammar (this study; Figure 7). *HOXA13* was initially detected in day 19 of the tammar forelimb, whilst in the mouse expression is first detected at E10.5 [6] and at stage 23 in the chicken [42]. There is a similar distal expression pattern in the tammar, chicken and mouse forelimb and the



expression has expanded proximally. At day 21, tammar *HOXA13* was weakly expressed and restricted to the interdigital regions. The expression boundary had shifted the anterior-posterior boundary but this pattern was similar to that of both the mouse and chicken. At

E11.5, mouse *HOXA13* expands in the proximal direction and a day later at E12.5 the first signs of weaker expression are detected in the digital condensations [43]. This is similar to the chicken that had expanded expression in the proximal direction by stage

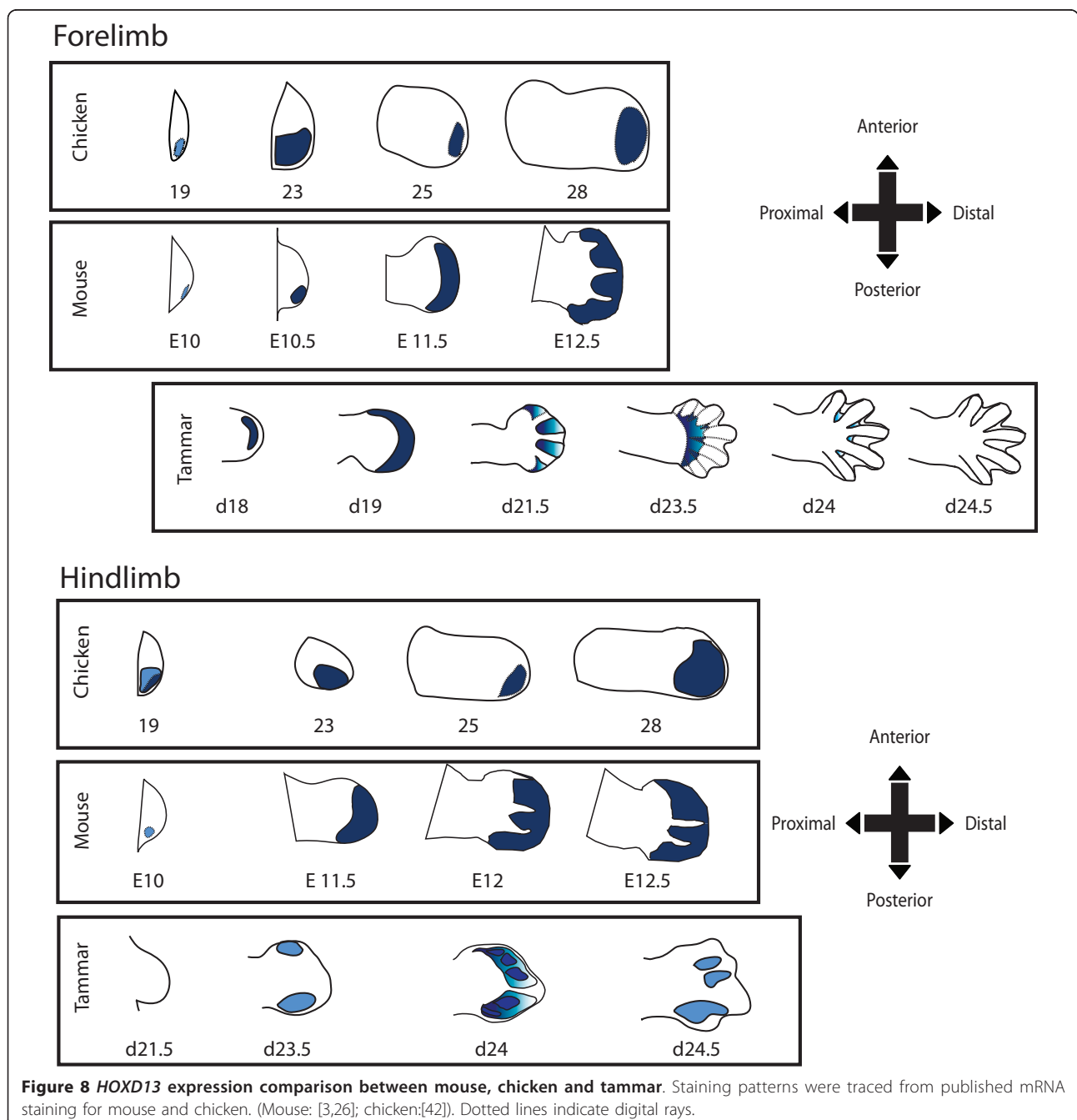
25 [42]. At stage 28, the chicken wing has strong posterior margins and expression is excluded in the region where the elongated digit of the forelimb will form.

There was a restricted distal expression pattern of *HOXA13* in the tammar hindlimb at day 21. In contrast, the chicken has a distal but expanded expression at stage 23 that was similar to the wing expression [42]. In contrast, the chicken leg at stage 25 has expanded expression towards the proximal axis and weakening expression in the distal region [42]. At stage 28 the

chicken leg has strong and expanded distal expression around the anterior and posterior margins. There was no detectable expression where the digital condensations form. The mouse has similar fore- and hind- limb expression of *HOXA13* [43] to the tammar and chicken forelimb.

Tammar, mouse and chicken *HOXD13* expression

HOXD13 expression patterns between tammar, mouse and chicken were also compared (Figure 8). At day 18 in



the tammar, *HOXD13* expression was first observed in the forelimb in the stages examined and there was strong distal expression that extended from the anterior and posterior ends of the paddle like structure. This expression was similar to that of the mouse at E11.5 forelimb [3] although the first detectable expression in the mouse occurs at E10.0 in early bud stage and at stage 18 in the chicken [42]. At day 21.5, tammar *HOXD13* became restricted to the interdigital regions, a pattern that was similar to the mouse E12.5 forelimb bud. *HOXD13* in the chicken wing is more distally restricted and unlike expression in the tammar and mouse does not extend from the most anterior point to the posterior margins [42]. At day 23.5, *HOXD13* in the tammar was less strongly expressed in the regions behind the digital condensations and in the interdigital regions. There is similar expression of *HOXD13* in the chicken and mouse as in the staining of the corresponding tammar forelimbs. At day 24 *HOXD13* was strongly expressed at the anterior and posterior margins with expression excluded where the digital condensations will form. A day later, one day before birth, there was no detectable expression.

Discussion

Tammar *HOXA13* and *HOXD13* genes are highly conserved with those of the chicken and mouse but there were unique expression patterns of *HOXA13* and *HOXD13* in the developing limbs. In particular, the syndactylous digits of the hindlimb began their differentiation pre-natally, but there were marked differences in the time of expression of these genes in both the fore- and hindlimbs, supporting the suggestion that the HOX genes are as important for patterning of the marsupial autopod as they are in other mammals.

The tammar limb shows heterochrony

The tammar forelimb is well advanced in the relative time of development compared to the mouse and chicken. At all stages, the development of the forelimb appears to be approximately two stages ahead of the hindlimb. This shift in developmental timing (heterochrony) is seen in other marsupial species like the opossum and the dasyurids [31]. However, the tammar has the most developed neonate amongst marsupials (Grade 3 as defined by Hughes and Hall, 1988) and has especially well-defined forelimbs that it uses to climb to the pouch. The control of heterochrony in marsupials remains unknown and an empirical analysis of tammar timing is not examined here, but in the opossum there is greater forelimb myocyte allocation compared with that of the mouse [29]. This heterochronic shift is a two-fold process with an acceleration of the forelimb and a delay in the development of the hindlimb bud

[31]. However, post-natally, there is a rapid catch-up growth in the tammar hindlimb.

The syndactylous digits are distinct in early development

The tammar hindlimb digits show clear evidence of syndactyly before birth. The rays of digits two and three are visible before any bone has formed in the hindlimbs. The early tammar fore and hindlimb buds both progress through a paddle-like stage, but the hindlimb soon becomes asymmetric. An early change in bud morphology is in accord with other species that also have reduced digits such as the chicken [42]. The tammar precursors of pedal digit 1 are not present at any stage, and the tammar hindlimb bud is more asymmetric than the chicken that has lost 2 digits. A reduction in the allocation of cells destined for the condensation of the missing digit may explain its loss [44]. Across different dog breeds there is variability in digit number and the number of digits may be due to the variability of the size of the breed and the number of cells in the embryonic limb [44].

Marsupial *HOXA13* and *HOXD13* are highly conserved

Tammar hindlimbs are reminiscent of mice and humans that have *HOXA13* and *HOXD13* mutations. However, the marsupial gene structure is highly conserved with mouse and opossum except for the N-terminal region of *HOXD13*. As expected, tammar *HOXA13* and *HOXD13* are highly conserved within all major vertebrate groups analysed, a phylogenetic tree (see Additional file 2) constructed with the predicted amino acids and others retrieved from NCBI (<http://www.ncbi.nlm.nih.gov/protein>) or Ensembl (<http://asia.ensembl.org/index.html>) produced tight branches. Given the high conservation of gene structure the specialised marsupial hindlimb is likely to be influenced through modification of gene expression domains, and timing or changes to downstream targets.

In mice lacking *HOXA13*, there are two potential downstream targets, *Gdf5* and *Bmp2* [45]. *Gdf5* and *Bmp2* are up- and down- regulated respectively and have diffuse expression in the carpal and tarsal anlagen. Interestingly, these mice also have a range of phenotypes including a fusion of carpal or tarsal elements and an absence of digit 1 in the hindlimb [16,45], a phenotype somewhat similar to that of the tammar hindlimb. *HOXA13* is also expressed at an equal level in all digits of the limb including digit 1. Macropodid marsupial hindlimbs lack digit 1 and have fused tarsals, so by analogy *HOXA13* is likely to have been involved in the evolution of these kangaroo phenotypes.

Determining the regulatory control of HOX genes is crucial in understanding the evolution of the different digital forms. The regulation of the HOX cluster is

believed to be controlled by a quantitative mechanism involving both gene topography and dosage. The relative proximity of *HOX* genes to two enhancers PROX and GCR located on the 5' end determines how strongly *HOX* genes are expressed [46]. In mice *HOXD13* is located towards the 5' end of the cluster and is expressed lower in the region destined to become digit one. In contrast, in the tammar forelimb which has a digit 1, we did not detect a difference in expression of *HOXD13* in any digit. Interestingly, in the hindlimb with its developing syndactylous digits, there is a lack of expression in the region of the prospective digit 4. This is similar to the situation observed in the bat and chick where 5'*HOXD* expression is restricted to the interdigital regions and excluded from condensing mesenchyme [26,47]. In addition, *HOXD* expression is believed to play a role in determining digit size and number through a dose dependent mechanism [48].

Interestingly, there has been at least one report of polydactyly in the forelimb of a kangaroo [49], a phenotype reminiscent of mutations in human *HOXA13* and *HOXD13*. Expansions or deletion in the polyalanine tracts of *HOXA13* and *HOXD13* is associated with these mutant phenotypes in man and mouse [50]. However, in dogs the variation in the number of tandem repeats in the genome is correlated with changes in limb and skull form [51]. In particular, the change in repeat length in *Aristaless-like 4 (ALX4)* observed between different dog species, was associated with formation of a rear first polydactyl digit [51].

Other vertebrate groups such as cetaceans have a novel expansion of the polyalanine tract in *HOXD13* compared with humans and mice, indicating it may influence the morphological diversity of the cetacean autopod [52]. Chicken and *Zebrafish* polyalanine tracts are shorter and less frequent compared to those of mammals. However, most polyalanine mutations lead to protein mis-folding, degradation and cytoplasmic aggregation and can repress expression depending on the number of polyalanines [19,50]. Bats have highly modified forelimbs with a greatly elongated third digit and have shifted the *HOXD13* anterior-posterior limits in the forelimb compared to the mouse but retain conserved polyalanine tracts [50]. The tammar has also shifted the *HOXD13* anterior-posterior boundaries (in the hindlimb), but does not have any mutation in the first and third polyalanine tracts (Figure 4) and so this cannot account for the unique tammar hindlimb. However, the first polyserine tract was missing in the N-terminal of *HOXD13* in both tammar and opossum, and instead possessed a unique series of amino acids in the tammar. It is possible that these six amino acids could affect the limb phenotype. The secondary structure of *HOXD13* in the tammar included a long 13 α -helix, but

in the human and the opossum it was only a 3 α -helix (Figure 4), suggesting that this region may be important in the development of a macropodid-specific form.

Conclusions

This study is the first to describe *HOX* expression in any marsupial. The subtle differences in gene structure in the tammar and the changes in expression and timing may drive the differences in the development of the syndactylous limb. *HOXA13* and *HOXD13* gene structures are highly conserved between marsupial, chicken and mouse. The lack of polyalanine modifications suggests these regions in either gene are unlikely to be the cause of altered limb morphology in the tammar but that the polyserine region may well be responsible for the development of marsupial syndactyly. Our findings support the hypothesis that changes to the structure and function of *HOXA13* and *HOXD13* affect regulation of digit identity in this marsupial.

Additional material

Additional file 1: Table S1: Protein sequences of *HOXA13* and *HOXD13*. The sequences used in this study were retrieved from GenBank or Ensembl.

Additional file 2: Phylogenetic tree of *HOXA13* and *HOXD13*. The evolutionary history was inferred using the Neighbor-Joining method [56]. The bootstrap consensus tree inferred from 1000 replicates is taken to represent the evolutionary history of the taxa analyzed [57]. Branches corresponding to partitions reproduced in less than 50% bootstrap replicates were collapsed. The percentage of replicate trees in which the associated taxa clustered together in the bootstrap test (1000 replicates) are shown next to the branches [57]. The evolutionary distances were computed using the Poisson correction method [37] and are in the units of the number of amino acid substitutions per site. The analysis involved 24 amino acid sequences with a total of 114 positions in the final dataset. All positions containing gaps and missing data were eliminated. Evolutionary analyses were conducted in MEGA5 [37].

Acknowledgements

We thank the member of the tammar research team for assistance of animal maintenance and collection. We also thank the three anonymous reviewers that made some very useful suggestion that helped to improve this manuscript. This study was supported by the Australian Research council (ARC) Centre of excellence in Kangaroo Genomics; an ARC Federation Fellowship to MBR and a National Health and Medical Research Council R.D Wright Fellowship to AJP.

Author details

¹ARC Centre of Excellence in Kangaroo Genomics, The University of Melbourne, Melbourne, Victoria, 3010, Australia. ²Department of Zoology, The University of Melbourne, Melbourne, Victoria, 3010, Australia. ³Department of Molecular and Cellular Biology, The University of Connecticut, Storrs, CT 06269, USA.

Authors' contributions

The authors declare the following contributions: conceived the study and design of experiments: KYC, HY and MBR. Collected tissues: KYC, HY, GS and MBR. Performed the experiments and analysis: KYC, HY and MBR. Manuscript preparation and discussion: KYC, HY, AJP and MBR. All authors read and approved the final manuscript.

Received: 3 August 2011 Accepted: 11 January 2012
Published: 11 January 2012

References

1. Sears KE: Novel insights into the regulation of limb development from 'natural' mammalian mutants. *BioEssays* 2011, 1-5.
2. Niswander L: Pattern formation: Old models out on a limb. *Nature Reviews Genetics* 2003, **4**(2):133-143.
3. Keyte A, Smith K: Opossum (monodelphis domestica): A marsupial development model. *Cold Spring Harbour Protocols* 2008, **2008**:104.
4. Cretekos CJ, Weatherbee SD, Chen CH, Badwaik NK, Niswander L, Behringer RR, Rasweiler I, John J: Embryonic staging system for the short tailed fruit bat, carollia perspicillata, a model organism for the mammalian order chiroptera, based upon timed pregnancies in captive bred animals. *Developmental Dynamics* 2005, **233**(3):721-738.
5. Wood Jones F: *The mammals of south australia* Adelaide, Australia: Government Printer; 1925.
6. Zákány J, Duboule D: Hox genes in digit development and evolution. *Cell and Tissue Research* 1999, **296**(1):19-25.
7. Lewandoski M, Sun X, Martin GR: Fgf8 signalling from the aer is essential for normal limb development. *Nature genetics* 2000, **26**(4):460-463.
8. Weatherbee SD, Behringer RR, Rasweiler JJ, Niswander LA: Interdigital webbing retention in bat wings illustrates genetic changes underlying amniote limb diversification. *Proceedings of the National Academy of Sciences of the United States of America* 2006, **103**(41):15103-15107.
9. Bastida MF, Sheth R, Ros MA: A bmp-shh negative-feedback loop restricts shh expression during limb development. *Development* 2009, **136**(22):3779.
10. Cohn MJ: Development of the external genitalia: Conserved and divergent mechanisms of appendage patterning. *Developmental Dynamics* 2011, **240**(5):1108-1115.
11. Yamada G, Suzuki K, Haraguchi R, Miyagawa S, Satoh Y, Kamimura M, Nakagata N, Kataoka H, Kuroiwa A, Chen Y: Molecular genetic cascades for external genitalia formation: An emerging organogenesis program. *Developmental Dynamics* 2006, **235**(7):1738-1752.
12. Miyagawa S, Matsumaru D, Murahisa A, Omori A, Satoh Y, Haraguchi R, Motoyama J, Iguchi T, Nakagata N, Hui C: The role of sonic hedgehog-gli2 pathway in the masculinization of external genitalia. *Endocrinology* 2011, **152**(7):2894.
13. Deschamps J, van Nes J: Developmental regulation of the hox genes during axial morphogenesis in the mouse. *Development* 2005, **132**(13):2931-2942.
14. Zakany J, Duboule D: The role of hox genes during vertebrate limb development. *Current Opinion in Genetics & Development* 2007, **17**(4):359-366.
15. Krumlauf R: Hox genes in vertebrate development. *Cell* 1994, **78**(2):191-201.
16. Fromental-Rainain C, Warot X, Messadecq N, LeMeur M, Dolle P, Chambon P: Hoxa-13 and hoxd-13 play a crucial role in the patterning of the limb autopod. *Development* 1996, **122**(10):2997-3011.
17. Frisén L: *Genetic studies of hypospadias* 2002.
18. Bruneau S, Johnson KR, Yamamoto M, Kuroiwa A, Duboule D: The mouse hoxd13(spdh) mutation, a polyalanine expansion similar to human type ii synpolydactyly (spd), disrupts the function but not the expression of other hoxd genes. *Developmental biology* 2001, **237**(2):345-353.
19. Amiel J, Trochet D, Clement-Ziza M, Munnich A, Lyonnet S: Polyalanine expansions in human. *Human Molecular Genetics* 2004, **13**(Review Issue 2):235.
20. Debeer P, Bacchelli C, Scambler P, De Smet L, Fryns J, Goodman F: Severe digital abnormalities in a patient heterozygous for both a novel missense mutation in hoxd13 and a polyalanine tract expansion in hoxa13. *Journal of medical genetics* 2002, **39**(11):852.
21. Innis JW, Mortlock D, Chen Z, Ludwig M, Williams ME, Williams TM, Doyle CD, Shao Z, Glynn M, Mikulic D: Polyalanine expansion in hoxa13: Three new affected families and the molecular consequences in a mouse model. *Human Molecular Genetics* 2004, **13**(22):2841.
22. Goodman FR: Limb malformations and the human hox genes. *American journal of medical genetics* 2002, **112**(3):256-265.
23. Lavoie H, Debeane F, Trinh QD, Turcotte JF, Corbeil-Girard LP, Dicaire MJ, Saint-Denis A, Pagé M, Rouleau GA, Brais B: Polymorphism, shared functions and convergent evolution of genes with sequences coding for polyalanine domains. *Human Molecular Genetics* 2003, **12**(22):2967.
24. Kohlhaase J, Wischermann A, Reichenbach H, Froster U, Engel W: Mutations in the sall1 putative transcription factor gene cause townes-brocks syndrome. *Nature genetics* 1998, **18**(1):81-83.
25. Kawakami Y, Uchiyama Y, Rodriguez Esteban C, Inenaga T, Koyano-Nakagawa N, Kawakami H, Marti M, Kmita M, Monaghan-Nichols P, Nishinakamura R, Izpisua Belmonte JC: Sall genes regulate region-specific morphogenesis in the mouse limb by modulating hox activities. *Development* 2009, **136**(4):585-594.
26. Chen C-H, Cretekos CJ, Rasweiler JJ, Behringer RR: Hoxd13 expression in the developing limbs of the short-tailed fruit bat, carollia perspicillata. *Evolution & Development* 2005, **7**(2):130-141.
27. Hockman D, Cretekos CJ, Mason MK, Behringer RR, Jacobs DS, Illing N: A second wave of sonic hedgehog expression during the development of the bat limb. *Proceedings of the National Academy of Sciences* 2008, **105**(44):16982.
28. Keyte A, Imam T, Smith K: Limb heterochrony in a marsupial, m. domestica. *Developmental biology* 2006, **295**(1):415.
29. Keyte AL, Smith KK: Developmental origins of precocial forelimbs in marsupial neonates. *Development* 2010, **137**(24):4283.
30. Bininda-Emonds ORP, Jeffery JE, Sánchez-Villagra MR, Hanken J, Colbert M, Pieau C, Selwood L, Ten Cate C, Raynaud A, Osabutey CK: Forelimb-hindlimb developmental timing changes across tetrapod phylogeny. *BMC Evolutionary Biology* 2007, **7**(1):182.
31. Sears KE: Differences in the timing of prechondrogenic limb development in mammals: The marsupial-placental dichotomy resolved. *Evolution* 2009, **63**(8):2193-2200.
32. McCrady E: *The embryology of the opossum* 1938, The Wistar institute of anatomy.
33. Manipulation of marsupial embryos and pouch young. In *Methods in mammalian reproduction* Edited by: Renfree M, Tyndale-Biscoe C, J Daniel Jr 1978, 307-331.
34. Renfree MB, Tyndale-Biscoe C: Intrauterine development after diapause in the marsupial macropus eugenii. *Developmental biology* 1973, **32**(1):28-40.
35. Tyndale-Biscoe CH, Renfree M: *Reproductive physiology of marsupials* Cambridge Univ Press; 1987.
36. Edgar RC: Muscle: Multiple sequence alignment with high accuracy and high throughput. *Nucleic acids research* 2004, **32**(5):1792.
37. Zuckerkandl E, Pauling L: Evolutionary divergence and convergence in proteins. *Evolving genes and proteins* 1965, **97**:166.
38. Smith CA, Smith MJ, Sinclair AH: Expression of chicken steroidogenic factor-1 during gonadal sex differentiation. *General and Comparative Endocrinology* 1999, **113**(2):187-196.
39. Nagy A, Gertsenstein M, Vintersten K, Behringer R: Alcian blue staining of the mouse fetal cartilaginous skeleton. *Cold Spring Harbor Protocols* 2009, **2009**(3), pdb.prot5169.
40. Nelson CE, Morgan BA, Burke AC, Laufer E, DiMambro E, Murtaugh LC, Gonzales E, Tessarollo L, Parada LF, Tabin C: Analysis of hox gene expression in the chick limb bud. *Development* 1996, **122**(5):1449-1466.
41. Salsi V, Zappavigna V: Hoxd13 and hoxa13 directly control the expression of the epha7 ephrin tyrosine kinase receptor in developing limbs. *Journal of Biological Chemistry* 2006, **281**(4):1992-1999.
42. Nelson CE, Morgan B, Burke A, Laufer E, DiMambro E, Murtaugh L, Gonzales E, Tessarollo L, Parada L, Tabin C: Analysis of hox gene expression in the chick limb bud. *Development* 1996, **122**(5):1449.
43. Albrecht AN, Schwabe GC, Stricker S, Böddrich A, Wanker EE, Mundlos S: The synpolydactyly homolog (spdh) mutation in the mouse - a defect in patterning and growth of limb cartilage elements. *Mechanisms of Development* 2002, **112**(1-2):53-67.
44. Alberch P: Developmental constraints: Why st. Bernards often have an extra digit and poodles never do. *The American Naturalist* 1985, **126**(3):430-433.
45. Perez WD, Weller CR, Shou S, Stadler HS: Survival of hoxa13 homozygous mutants reveals a novel role in digit patterning and appendicular skeletal development. *Developmental Dynamics* 2010, **239**(2):446-457.
46. Montavon T, Le Garrec J-F, Kerszberg M, Duboule D: Modeling hox gene regulation in digits: Reverse collinearity and the molecular origin of thumbness. *Genes & development* 2008, **22**(3):346-359.
47. Carkett M, Logan M: 1, 2, 3: Counting the fingers on a chicken wing. *Genome Biology* 2011, **12**(10):130.

48. Zákány J, Fromental-Ramain C, Warot X, Duboule D: **Regulation of number and size of digits by posterior hox genes: A dose-dependent mechanism with potential evolutionary implications.** *Proceedings of the National Academy of Sciences* 1997, **94**(25):13695-13700.
49. de Paz F, Pastor J, Gil J, Barbosa M, de la Iglesia M, Garcia M: **A case of bilateral polydactyly in the grey kangaroo (*macropus giganteus*).** *EUROPEAN JOURNAL OF ANATOMY* 2007, **11**(3):189..
50. Brown LY, Brown SA: **Alanine tracts: The expanding story of human illness and trinucleotide repeats.** *Trends in Genetics* 2004, **20**(1):51-58.
51. Fondon JW, Garner HR: **Molecular origins of rapid and continuous morphological evolution.** *Proceedings of the National Academy of Sciences of the United States of America* 2004, **101**(52):18058.
52. Wang Z, Yuan L, Rossiter SJ, Zuo X, Ru B, Zhong H, Han N, Jones G, Jepson PD, Zhang S: **Adaptive evolution of 5 *hoxd* genes in the origin and diversification of the cetacean flipper.** *Molecular biology and evolution* 2009, **26**(3):613.
53. Hume Id, Marilyn PJJ, Renfree B, Peter D: **Temple-Smith, *Macropodidae*, in Fauna of australia mammalia.** Australian Government Publishing service: Canberra; 1989, 679-715.
54. Zúñiga A, Zeller R: **Gli3 (xt) and formin (ld) participate in the positioning of the polarising region and control of posterior limb-bud identity.** *Development* 1999, **126**(1):13.
55. Bruneau S, Johnson KR, Yamamoto M, Kuroiwa A, Duboule D: **The mouse *hoxd13^{spd}* mutation, a polyalanine expansion similar to human type ii synpolydactyly (*spd*), disrupts the function but not the expression of other *hoxd* genes.** *Developmental biology* 2001, **237**(2):345-353.
56. Saitou N, Nei M: **The neighbor-joining method: A new method for reconstructing phylogenetic trees.** *Molecular biology and evolution* 1987, **4**(4):406..
57. Felsenstein J: **Confidence limits on phylogenies: An approach using the bootstrap.** *Evolution* 1985, 783-791.

doi:10.1186/1471-213X-12-2

Cite this article as: Chew *et al.*: *HOXA13* and *HOXD13* expression during development of the syndactylous digits in the marsupial *Macropus eugenii*. *BMC Developmental Biology* 2012 **12**:2.

**Submit your next manuscript to BioMed Central
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

